

Mouse SCF ELISA Kit

Vertrieb:

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Hinweis/Note:

Der Packungsbeileger dient nur als erste Information. Der relevante Packungsbeileger liegt der Ware bei.

The datasheet is only a first information.

The relevant datasheet is included with the product.

For any questions regarding troubleshooting or performing the assay, please contact our support team at support@assaypro.com.

Thank you for choosing Assaypro.

Assay Summary

Add 50 μl of Standard/ Sample per well. Incubate 2 hours.



Wash, then add 50 μl of Biotinylated Antibody per well. Incubate 1 hour.



Wash, then add 50 μl of SP Conjugate per well. Incubate 30 minutes.



Wash, then add 50 μl of Chromogen Substrate per well. Incubate 15 minutes.



Add 50 µl of Stop Solution per well. Read at 450 nm immediately.

Assay Template

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AssayMax Mouse Stem Cell Factor (SCF) ELISA Kit

Catalog No. EMS1001-1 Sample Insert/Reference Only

Introduction

Stem Cell Factor (SCF) is known as c-Kit receptor ligand, KL, steel factor, or mast cell growth factor and is expressed in fibroblasts, thymus tissue, spleen, testes, placenta, and mast cells. SCF is a cytokine that exists in two forms produced by alternative splicing: a soluble form of approximately 31 kDa and a membrane-bound form of approximately 32 kDa, lacking the proteolytic site for processing into the soluble form (1-4). SCF not only plays an important role in hematopoiesis, reproduction, melanogenesis, and tumor progression, but is also involved in proliferation and differentiation of mast cells. It stimulates mast cell activation in human bronchi and induces smooth muscle cell contraction (5). Both increased expression of SCF and its receptor c-Kit were found in asthma patients (6). During chronic stroke, SCF in combination with granulocyte-colony stimulating factor (G-CSF) treatment can enhance repair of brain damage (7). Blocking SCF-c-kit signaling is sufficient to inhibit lung cancer stem cell proliferation and survival promoted by chemotherapy (8).

Principle of the Assay

The AssayMax Mouse Stem Cell Factor ELISA (Enzyme-Linked Immunosorbent Assay) kit is designed for detection of mouse SCF in urine and cell culture samples. This assay employs a quantitative sandwich enzyme immunoassay technique that measures mouse SCF in less than 4 hours. A polyclonal antibody specific for mouse SCF has been pre-coated onto a 96-well microplate with removable strips. SCF in standards and samples is sandwiched by the immobilized antibody and the biotinylated polyclonal antibody specific for SCF, which is recognized by a streptavidin-peroxidase conjugate. All unbound material is then washed away and a peroxidase enzyme substrate is added. The color development is stopped and the intensity of the color is measured.

Caution and Warning

- Prepare all reagents (working diluent buffer, wash buffer, standard, biotinylated antibody, and SP conjugate) as instructed, prior to running the assay.
- Prepare all samples prior to running the assay. The dilution factors for the samples are suggested in this protocol. However, the user should determine the optimal dilution factor.
- Spin down the SP conjugate vial and the biotinylated antibody vial before opening and using contents.
- This kit is for research use only.
- The kit should not be used beyond the expiration date.
- The Stop Solution is an acidic solution.

Reagents

- **Mouse SCF Microplate:** A 96-well polystyrene microplate (12 strips of 8 wells) coated with a polyclonal antibody against mouse SCF.
- **Sealing Tapes:** Each kit contains 3 precut, pressure sensitive sealing tapes that can be cut to fit the format of the individual assay.
- Mouse SCF Standard: Mouse SCF in a buffered protein base (4 ng, lyophilized).
- **Biotinylated Mouse SCF Antibody (50x):** A 50-fold concentrated biotinylated polyclonal antibody against mouse SCF (140 μl).
- MIX Diluent Concentrate (10x): A 10-fold concentrated buffered protein base (30 ml).
- Wash Buffer Concentrate (20x): A 20-fold concentrated buffered surfactant (30 ml, 2 bottles).
- Streptavidin-Peroxidase Conjugate (SP Conjugate): A 100-fold concentrate (80 μl).
- **Chromogen Substrate**: A ready-to-use stabilized peroxidase chromogen substrate tetramethylbenzidine (8 ml).
- **Stop Solution**: A 0.5 N hydrochloric acid to stop the chromogen substrate reaction (12 ml).

Storage Condition

- Upon arrival, immediately store components of the kit at recommended temperatures up to the expiration date.
- Store SP Conjugate and Biotinylated Antibody at -20°C.
- Store Microplate, Diluent Concentrate (10x), Wash Buffer, Stop Solution, and Chromogen Substrate at 2-8°C.

- Unused microplate wells may be returned to the foil pouch with the desiccant packs and resealed. May be stored for up to 30 days in a vacuum desiccator.
- Diluent (1x) may be stored for up to 30 days at 2-8°C.
- Store Standard at 2-8°C before reconstituting with Diluent and at -20°C after reconstituting with Diluent.

Other Supplies Required

- Microplate reader capable of measuring absorbance at 450 nm.
- Pipettes (1-20 μl, 20-200 μl, 200-1000 μl, and multiple channel).
- Deionized or distilled reagent grade water.

Sample Collection, Preparation and Storage

- **Cell Culture Supernatants:** Centrifuge cell culture media at 3000 x g for 10 minutes to remove debris. Collect supernatants and assay. Store samples at -20°C or below. Avoid repeated freeze-thaw cycles.
- **Urine:** Collect urine using sample pot. Centrifuge samples at 800 x g for 10 minutes and assay. Store samples at -20°C or below for up to 3 months. Avoid repeated freeze-thaw cycles.

Reagent Preparation

- Freshly dilute all reagents and bring all reagents to room temperature before use.
- MIX Diluent Concentrate (10x): If crystals have formed in the concentrate, mix gently until the crystals have completely dissolved.
 Dilute the MIX Diluent Concentrate 1:10 with reagent grade water. Store for up to 30 days at 2-8°C.
- Standard Curve: Reconstitute the 4 ng of Mouse SCF Standard with 1 ml of MIX Diluent to generate a 4 ng/ml standard solution. Allow the standard to sit for 10 minutes with gentle agitation prior to making dilutions. Prepare duplicate or triplicate standard points by serially diluting the standard solution (4 ng/ml) 1:2 with MIX Diluent to produce 2, 1, 0.5, 0.25, 0.125, and 0.063 ng/ml solutions. MIX Diluent serves as the zero standard (0 ng/ml). Any remaining solution should be frozen at -20°C and used within 30 days.

Standard Point	Dilution	[Mouse SCF] (ng/ml)	
P1	Standard (4 ng/ml)	4.000	
P2	1 part P1 + 1 part MIX Diluent	2.000	
P3	1 part P2 + 1 part MIX Diluent	1.000	
P4	1 part P3 + 1 part MIX Diluent	0.500	
P5	1 part P4 + 1 part MIX Diluent	0.250	
P6	1 part P5 + 1 part MIX Diluent	0.125	
P7	1 part P6 + 1 part MIX Diluent	0.063	
P8	MIX Diluent	0.000	

- **Biotinylated Mouse SCF Antibody (50x):** Spin down the antibody briefly and dilute the desired amount of the antibody 1:50 with MIX Diluent. Any remaining solution should be frozen at -20°C.
- Wash Buffer Concentrate (20x): If crystals have formed in the concentrate, mix gently until the crystals have completely dissolved. Dilute the Wash Buffer Concentrate 1:20 with reagent grade water.
- **SP Conjugate (100x):** Spin down the SP Conjugate briefly and dilute the desired amount of the conjugate 1:100 with MIX Diluent. Any remaining solution should be frozen at -20°C.

Assay Procedure

- Prepare all reagents, standard solutions, and samples as instructed. Bring all reagents to room temperature before use. The assay is performed at room temperature (20-25°C).
- Remove excess microplate strips from the plate frame and return them immediately to the foil pouch with desiccants inside. Reseal the pouch securely to minimize exposure to water vapor and store in a vacuum desiccator.
- Add 50 μ l of Mouse SCF Standard or sample per well. Cover wells with a sealing tape and incubate for 2 hours. Start the timer after the last addition.
- Wash five times with 200 μ l of Wash Buffer manually. Invert the plate each time and decant the contents; hit 4-5 times on absorbent material to completely remove the liquid. If using a machine, wash six times with 300 μ l of Wash Buffer and then invert the plate, decanting the contents; hit 4-5 times on absorbent material to completely remove the liquid.
- Add 50 μ l of Biotinylated Mouse SCF Antibody to each well and incubate for 1 hour.
- Wash the microplate as described above.
- Add 50 μ l of Streptavidin-Peroxidase Conjugate to each well and incubate for 30 minutes. Turn on the microplate reader and set up the program in advance.
- Wash the microplate as described above.

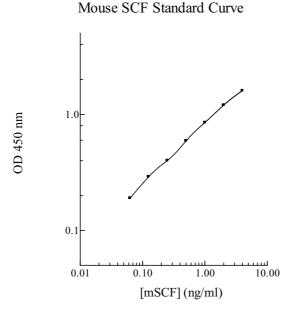
- Add 50 μ l of Chromogen Substrate per well and incubate for about 15 minutes or till the optimal blue color density develops. Gently tap plate to ensure thorough mixing and break the bubbles in the well with pipette tip.
- Add 50 μ l of Stop Solution to each well. The color will change from blue to vellow.
- Read the absorbance on a microplate reader at a wavelength of 450 nm immediately. If wavelength correction is available, subtract readings at 570 nm from those at 450 nm to correct optical imperfections. Otherwise, read the plate at 450 nm only. Please note that some unstable black particles may be generated at high concentration points after stopping the reaction for about 10 minutes, which will reduce the readings.

Data Analysis

- Calculate the mean value of the duplicate or triplicate readings for each standard and sample.
- To generate a standard curve, plot the graph using the standard concentrations on the x-axis and the corresponding mean 450 nm absorbance on the y-axis. The best-fit line can be determined by regression analysis using log-log or four-parameter logistic curve-fit.
- Determine the unknown sample concentration from the Standard Curve and multiply the value by the dilution factor.

Standard Curve

• The curve is provided for illustration only. A standard curve should be generated each time the assay is performed.



Performance Characteristics

- The minimum detectable dose of mouse SCF is typically less than ~ 0.06 ng/ml.
- Intra-assay and inter-assay coefficients of variation were 4.6% and 7.1% respectively.

Recovery

Standard Added Value	0.1 – 2 ng/ml
Recovery %	87 – 111%
Average Recovery %	96%

Cross-Reactivity

Species	% Cross Reactivity
Canine	5%
Bovine	None
Monkey	None
Human	None
Rat	None
Swine	5%
Rabbit	None
Mouse	100%

References

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